CORRESPONDENCE

MicroRNAs predict early complications of autologous hematopoietic stem cell transplantation

Damian Mikulski¹, Mateusz Nowicki^{2,3}, Izabela Dróżdż⁴, Ewelina Perdas¹, Piotr Strzałka^{2,3}, Kacper Kościelny¹, Małgorzata Misiewicz², Konrad Stawiski¹, Agnieszka Wierzbowska^{2,3} and Wojciech Fendler^{1,5*}

Abstract

Autologous hematopoietic stem cell transplantation (AHSCT) remains the most prevalent type of stem cell transplantation. In our study, we investigated the changes in circulating miRNAs in AHSCT recipients and their potential to predict early procedure-related complications. We collected serum samples from 77 patients, including 54 with multiple myeloma, at four key time points: before AHSCT, on the day of transplantation (day 0), and at days + 7 and + 14 post-transplantation. Through serum miRNA-seg analysis, we identified altered expression patterns and miRNAs associated with the AHSCT procedure. Validation using gPCR confirmed deviations in the levels of miRNAs at the beginning of the procedure in patients who subsequently developed bacteremia: hsa-miR-223-3p and hsa-miR-15b-5p exhibited decreased expression, while hsa-miR-126-5p had increased level. Then, a neural network model was constructed to use miRNA levels for the prediction of bacteremia. The model achieved an accuracy of 93.33% (95%Cl: 68.05-99.83%), with a sensitivity of 100% (95%Cl: 67.81-100.00%) and specificity of 90.91% (95%CI: 58.72-99.77%) in predicting bacteremia with mean of 6.5±3.2 days before occurrence. In addition, we showed unique patterns of miRNA expression in patients experiencing platelet engraftment delay which involved the downregulation of hsa-let-7f-5p and upregulation of hsa-miR-96-5p; and neutrophil engraftment delay which was associated with decreased levels of hsa-miR-125a-5p and hsa-miR-15b-5p. Our findings highlight the significant alterations in serum miRNA levels during AHSCT and suggest the clinical utility of miRNA expression patterns as potential biomarkers that could be harnessed to improve patient outcomes, particularly by predicting the risk of bacteremia during AHSCT.

Keywords miRNA, Autologous hematopoietic stem cell transplant, Hsa-miR-223-3p, Hsa-miR-15b-5p, Hsa-miR-126-5p, Bacteremia, Biomarkers

*Correspondence:

- Wojciech Fendler
- wojciech.fendler@umed.lodz.pl
- ¹Department of Biostatistics and Translational Medicine, Medical University of Lodz, Lodz, Poland

²Department of Hematology, Medical University of Lodz, Lodz, Poland

³Department of Hematology and Transplantology, Provincial Multi-

⁴Department of Clinical Genetics, Medical University of Lodz, Lodz, Poland

⁵Department of Radiation Oncology, Dana-Farber Cancer Institute, Boston, MA, USA



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.







Specialized Oncology and Trauma Center, Lodz, Poland

To the editor

Autologous hematopoietic stem cell transplantation (AHSCT) is broadly used to treat hematologic disorders (predominantly multiple myeloma), with an estimated 28,700 procedures performed in Europe in 2019 [1]. Attempts to establish AHSCT as an outpatient procedure are gaining traction, but concerns about adverse effects like mucositis, bacteremia or delayed engraftment (DE) limit this transition [2, 3]. Conventional cytokine or cell count-based biomarkers may be unreliable in predicting or detecting those complications in AHSCT recipients due to the nature of the procedure itself. In the present study, we aimed to quantify alterations in the signature of freely circulating miRNAs in the sera of AHSCT recipients and identify circulating miRNAs that could be used to create a predictive model for bacteremia - a common and potentially life-threatening complication of AHSCT [4-6].

Serum samples were taken from all patients (N=77; Table 1 and Supplementary Table 1) at four time points throughout AHSCT. miRNA-seq was performed to identify potential miRNA biomarkers (N1=10). MiRNAs with profiles affected by AHSCT were subsequently validated

 Table 1
 Clinical characteristics of patients included into the study

| Variable | N=77 (100%) | | | |
|---|------------------------|--------------|----------|--|
| Disease | MM· 54 (70.1) | | | |
| Discuse | HI · 11 (14 3) | | | |
| | MCL: 5 (6.5) | | | |
| | DLBCL: 3 (3.9) | | | |
| | Other: 4 (5.2) | | | |
| Conditioning regimen | Mel-200: 39 (50.6) | | | |
| | Reduced Mel: 16 (20.8) | | | |
| | BeEAM: 17 (22.1) | | | |
| | BEAM: 5 (6.5) | | | |
| Variable | MM | ML | р | |
| Sex | F: 29 (53.7) | F: 11 (47.8) | 0.8233 | |
| | M: 25 (46.3) | M: 12 | | |
| | | (52.2) | | |
| Age at AHSCT | 61.3 (53.8–66.4) | 47.4 | < 0.0001 | |
| Median (25–75%), years | | (39.5–54.1) | | |
| Number of CD34+cells | 3.9 (3.1–5.3) | 4.6 | 0.4092 | |
| transplanted (× 10 ⁶ cells/kg) | | (3.3–5.7) | | |
| Median (25–75%) | | | | |
| Mucositis- any grade | 34 (64.2) | 23 (100.0) | 0.0004 | |
| Mucositis– grade≥2 | 18 (34.0) | 22 (95.7) | < 0.0001 | |
| Bacteremia | 11 (20.4) | 8 (34.8) | 0.2920 | |
| Neutropenic fever | 30 (58.8) | 18 (78.3) | 0.1745 | |
| Days to PLT engraftment | 10 (10–13) | 15 (12–18) | 0.0001 | |
| Median (25–75%) | | | | |
| Days to ANC engraftment | 11 | 10 | 0.0299 | |
| Median (25–75%) | | | | |

ANC- absolute neutrophil count; BEAM- carmustine, etoposide, cytarabine, melphalan; BeEAM- bendamustine, etoposide, cytarabine, melphalan; DLBCLdiffuse large B-cell lymphoma; HL- Hodgkin Lymphoma; 25–75%- interquartile range; MCL- mantle cell lymphoma; MEL- melphalan; ML- malignant lymphoma; MM- multiple myeloma; PLT- platelets with a targeted qPCR (N2=67) for their association with bacteremia and other complications. The detailed Methods were presented in Supplementary File 1 and Supplementary Fig. 1.

In miRNA-seq data, dysregulation of miRNAs expression across study time points was shown with 20 miR-NAs showing a significant difference in global repeated measures ANOVA (Fig. 1A, Supplementary File 2). Twelve miRNAs were identified as eligible for qPCR validation (Fig. 1B and H and Supplementary Figs. 2 and 3) due to their significant fluctuations across the procedure and association with DE. Additional five miRNAs associated with irrevocable bone marrow damage (miR-150-5p, miR-375, miR-122-5p, hsa-miR-126-5p, and miR-122b-3p) and four potential reference miRNAs, two of which (hsa-miR-27b-3p and hsa-miR-148b-3p) were the final normalization factor to provide controls and calibration [7–9].

Overall, the results of both methods- miRNA-seq and RT-qPCR were highly convergent across all time points (Supplementary Fig. 4). All five miRNAs related to radio-therapy-induced response changed their expression significantly across the study time points (Supplementary Fig. 5) confirming their association with bone marrow damage.

In the RT-qPCR group, thirteen patients had neutrophil DE with lower expression of hsa-miR-125a-5p (FC_{T3}=0.77, p=0.0301) and hsa-miR-15b-5p (FC_{T1}=0.70, p=0.0428); while 13 had platelet DE time which was associated with hsa-let-7f-5p (FC_{T2}=0.59, p=0.0128) and hsa-miR-96-5p (FC_{T3}=1.82, p=0.0397) levels (Fig. 1I-J, Supplementary File 3).

In total, there were 17 episodes of documented bacteremia in the RT-qPCR set of patients. The majority were caused by Gram-positive bacteria (13, 76.5%). The mean time to bacteremia onset since AHSCT (T2) was 6.5 ± 3.2 days. Using miRNA levels at baseline or T2 a neural network (NN) model (Supplementary Files 4-7) for bacteremia prediction was iteratively designed. The final model relied on three miRNAs quantified at T2 were included: hsa-miR-223-3p, hsa-miR-15b-5p, and hsa-miR-126-5p (Fig. 1K, Supplementary Fig. 6) and showed accuracy of 93.33%, 95%CI:68.05-99.83% (Supplementary File 6) in the validation group with one false positive case occurred in the validation set (sensitivity 100%; specificity 90.91%, 95%CI: 58.72-99.77%). With the prevalence of bacteremia in the entire studied group, the positive predictive value reached 94.12% (95%CI: 69.61-99.11%) while NPV equaled 98.00% (95%CI: 87.97-99.70%).

Pathway analysis using the KEGG database demonstrated that miRNAs retained in the model were enriched for genes associated with various infections and responses to infections, including Hepatitis C, Toxoplasmosis, Salmonella infection, Shigellosis, Influenza A,



Fig. 1 (See legend on next page.)

(See figure on previous page.)

Fig. 1 miRNA-seq analysis results. Samples were drawn at four timepoints: (T1) before conditioning chemotherapy, (T2) on the day of AHSCT (day 0), day +7 (T3), and +14 day after AHSCT (T4). (**A**) heatmap of miRNAs differently expressed across study timepoint assessed by repeated measures ANOVA. The serum miRNA profiles tend to cluster by the study time points- two clusters- "early" (T1 and T2) and "late" (T3 and T4) are visible. One minus Pearson correlation distance metric and complete linkage method were used. (**B-H**) Plots for seven miRNAs differentially expressed across AHSCT procedure in miRNA-seq stage of the study. There were no statistically significant results in the comparison of miRNAs expression between at T1 and T2. In T3, hsa-miR-320c (**B**) was significantly upregulated compared to T1 (FC=3.92, p=0.007), whereas hsa-miR-223-3p (**C**) was significantly downregulated (FC 0.31, p=0.048). MiRNA levels at T4, in comparison to T1, showed significant downregulation of both hsa-let-7f-5p (**D**) and hsa-miR-155-5p (**E**) (FC=0.38, p=0.004 and FC=0.37, p=0.019, respectively), while hsa-miR-320c (**B**) was significantly upregulated (FC =2.39, p=0.049). At T3, there was lower expression level of hsa-miR-18a-5p (**F**) (FC=0.16, p=0.035) and hsa-miR-223-3p (**C**) (FC=0.35, p=0.033) comparing to T2. Comparing T4 with T3, a lower expression level of hsa-miR-486-5p (**G**) (FC=0.72, p=0.024) was identified. In a comparison of T4 with T2, a higher expression level of hsa-miR-96-5p (**H**) (FC=8.33, p=0.036) was observed. Asterisks denote the significance level (paired t-test with Bonferroni correction): *- $p \le 0.05$; **- $p \le 0.01$. (**I-J**) Volcano plots showing differentially expressed miRNAs in patients with platelet delayed engraftment (DE) (**I**) and neutrophil DE (**J**). Red dots represent upregulated miRNAs; blue dots represent downregulated miRNAs; grey dots represent miRNAs with no significant difference. (**K**) Volcano plot showing differentially expressed miRNAs in patients with platelet delayed en

Measles, Herpes simplex infection, Bacterial invasion of epithelial cells and Fc gamma R-mediated phagocytosis (Supplementary Figs. 7 and 8). Moreover, all three miR-NAs included in our model were identified and predicted to originate from potential tissue sources that are predominantly affected by the AHSCT procedure (Supplementary Fig. 9).

Our study is the first to assess circulating miRNA expression patterns during AHSCT and identify biomarkers of the procedure's complications. Notably, we observed expression changes in relation to complications such as bacteremia and engraftment delay. Interestingly, the differentially expressed miRNAs largely manifested prior to the onset of these complications. Our findings culminated in developing a predictive model distinguishing patients at risk of developing bacteremia– a critical and life-threatening AHSCT complication [4].

While we strove to include a balance of different indications for AHSCT, the relatively small sample size may have resulted in a bias toward the variable selection of miRNAs associated with particular underlying diseases. Replicable patterns of miRNAs identified earlier as associated with bone marrow damage seem to show that severe stimuli exert expression changes that are evident despite baseline differences [7, 10]. The evidence suggests that the individual miRNAs integrated into our model have also been independently associated with sepsis and severe infections across diverse patient cohorts. Specifically, hsa-miR-223-3p, hsa-miR-15b-5p, and hsa-miR-126-5p have consistently demonstrated connections to these events, irrespective of the underlying hematologic diagnoses [11, 12]. Nevertheless, independent external validation would strengthen their clinical relevance. Our findings regarding the association of circulating miRNA expression patterns incurred by bone marrow damage could extend beyond the setting of AHSCT, aiding targeted interventions to mitigate myelotoxicity and enhance the safety of other cancer treatments or detection of exposure to myelotoxic stimuli.

The underlying prior data on miRNA biomarkers of myelotoxicity concerned the Total Body Irradiation (TBI) procedure [7, 8]. In those patients - with different malignancies, clinical factors, and procedures- the impact of miRNAs was clearly evidenced and maintained regardless of clinical confounding factors. We thus hypothesize that the myeloablative procedure is an event of such catastrophic impact on the organism level that it overshadows other causes of miRNA expression variability at the serum level. Deregulated miRNAs consistently changed post high versus low radiation doses, with hsamiR-150-5p, hsa-miR-122-5p, hsa-miR-122b-3p decreasing, and hsa-miR-375, hsa-miR-126-5p increasing after radiotherapy [7]. In the current study, hsa-miR-150-5p declined, while hsa-miR-375 and hsa-miR-126-5p were over-expressed across AHSCT, mirroring changes during TBL.

In conclusion, our study shows distinct patterns of miRNA in chemotherapy-induced injury across AHSCT which may be used to predict bacteremia and potentially stratifying patients as eligible for outpatient AHSCT.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s40364-024-00585-x.

| Supplementary Material 1 | |
|--------------------------|--|
| Supplementary Material 2 | |
| Supplementary Material 3 | |
| Supplementary Material 4 | |
| Supplementary Material 5 | |
| Supplementary Material 6 | |
| Supplementary Material 7 | |

Acknowledgements

We thank all the patients who participated in the study and staff from the Department of Hematology and Transplantology, Copernicus Memorial Hospital in Lodz, Poland.

Author contributions

DM wrote the first version of the manuscript. DM and WF designed and planned the experiments. DM, MN, MM, PS, KK collected the data. DM, MN, MM and PS collected the samples. DM and IZ performed the experiments. DM, KS and WF performed statistical analyses. All authors reviewed and approved the submitted version.

Funding

The study was funded by the National Science Center grants number 2019/33/B/NZ5/00536 awarded to W.F. and 2022/45/N/NZ6/02904 awarded to DM.

Data availability

All data generated and analyzed during this study are included in this published article and its supplementary files.

Declarations

Ethics approval and consent to participate

Each patient signed the informed consent for all examinations and procedures. All procedures were approved by the local ethical committee (The Ethical Committee of the Medical University of Lodz, No RNN/424/19/KE).

Consent for publication

Not applicable.

Competing interests

A.W.: research grants: Jazz Pharmaceuticals; honoraria: AbbVie, Astellas, BMS/ Celgene, Gilead/Kite, Janssen, Novartis, Pfizer, Servier; advisory boards: AbbVie, Astellas, BerGenBio, BMS/Celgene, Gilead/Kite, Janssen, Novartis, Pfizer, Servier. The remaining authors have no conflicts of interest to declare.

Received: 13 October 2023 / Accepted: 28 March 2024 Published online: 23 April 2024

References

 Baldomero H, Passweg J. 30 Years EBMT ACTIVITY SURVEY and 2019 annual report [Internet]. https://www.ebmt.org/sites/default/files/2021-03/Transplant Activity Survey.pdf.

- Shah N, Cornelison AM, Saliba R, Ahmed S, Nieto YL, Bashir Q, et al. Inpatient vs outpatient autologous hematopoietic stem cell transplantation for multiple myeloma. Eur J Haematol. 2017;99(6):532–5.
- 3. Lutfi F, Skelton IVWP, Wang Y, Rosenau E, Farhadfar N, Murthy H, et al. Clinical predictors of delayed engraftment in autologous hematopoietic cell transplant recipients. Hematol Oncol Stem Cell Ther. 2020;13(1):23–31.
- Poutsiaka DD, Price LL, Ucuzian A, Chan GW, Miller KB, Snydman DR. Blood stream infection after hematopoietic stem cell transplantation is associated with increased mortality. Bone Marrow Transpl. 2007;40(1):63–70.
- Tomasik B, Papis-Ubych A, Stawiski K, Fijuth J, Kędzierawski P, Sadowski J, et al. Serum MicroRNAs as Xerostomia biomarkers in patients with Oropharyngeal Cancer Undergoing Radiation Therapy. Int J Radiat Oncol Biol Phys. 2021;111(5):1237–49.
- Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable blood-based markers for cancer detection. Proc Natl Acad Sci U S A. 2008;105(30):10513–8.
- Nowicka Z, Tomasik B, Kozono D, Stawiski K, Johnson T, Haas-Kogan D et al. Serum miRNA-based signature indicates radiation exposure and dose in humans: a multicenter diagnostic biomarker study. Radiother Oncol. 2023.
- Acharya SS, Fendler W, Watson J, Hamilton A, Pan Y, Gaudiano E, et al. Serum microRNAs are early indicators of survival after radiation-induced hematopoietic injury. Sci Transl Med. 2015;7(287):ra28769–28769.
- Grabia S, Smyczynska U, Pagacz K, Fendler W. NormiRazor: tool applying GPUaccelerated computing for determination of internal references in microRNA transcription studies. BMC Bioinformatics. 2020;21(1):425.
- Fendler W, Malachowska B, Meghani K, Konstantinopoulos PA, Guha C, Singh VK et al. Evolutionarily conserved serum microRNAs predict radiationinduced fatality in nonhuman primates. Sci Transl Med. 2017;9(379).
- Liu D, Wang Z, Wang H, Ren F, Li Y, Zou S et al. The protective role of miR-223 in sepsis-induced mortality. Scientific Reports. 2020 10:1 [Internet]. 2020 Oct 19 [cited 2024 Mar 13];10(1):1–10. https://www.nature.com/articles/ s41598-020-74965-2
- Goodwin AJ, Guo C, Cook JA, Wolf B, Halushka PV, Fan H. Plasma levels of microRNA are altered with the development of shock in human sepsis: an observational study. Crit Care [Internet]. 2015 Dec 18 [cited 2024 Mar 13];19(1). https://pubmed.ncbi.nlm.nih.gov/26683209/

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.