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The importance of critically short telomere in myelodysplastic syndrome



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Abstract

A few critically short telomeres trigger genomic instability regardless of average telomere length (TL). Recently, the telomere shortest length assay (TeSLA) was developed to detect critically short telomeres and measure absolute telomeres. Using TeSLA with the internally labeled biotin probe, we measured the TL of bone marrow (BM) aspirates from 52 patients with myelodysplastic syndrome (MDS). A percentage of shortest telomeres (<1.0 kb (ShTL1.0)) were calculated. ShTL1.0 was correlated to IPSS-R risk (spearman's rho = 0.35 and p = 0.0196), and ShTL1.0 and BM blast (2.61% in < 5% blast, 4.15% in 5–10% blast, and 6.80% in 10–20% blast, respectively, p = 0.0332). Interestingly, MDS patients with a shortest TL \geq 0.787 kb at the time of diagnosis showed better overall survival (OS) and progression-free survival (PFS) than patients with a shortest TL < 0.787 kb in the multivariate analyses (HR = 0.13 and 0.30, p = 0.011 and 0.048 for OS and PFS, respectively). Our results clearly show the presence and abundance of critically short telomeres in MDS patients. These pathologic telomeres are associated with IPSS-R which is a validated prognostic scoring system in MDS. Furthermore, they are independent prognostic factors for OS in MDS patients. Future prospective studies are needed to validate our results.

Highlights

Telomere length (TL) has been reported to be important in myelodysplastic syndrome (MDS). A novel TeSLA method demonstrated the presence and abundance of extremely short telomeres (<1.0kb) in MDS.Critically short TL rather than an average TL is associated with the IPSS-R and BM blast in MDS.The shortest TL is an independent prognostic factor for PFS and OS.Short TL should be incorporated into the risk scoring system in MDS in the future.

Keywords: TeSLA, Short telomere, Myelodysplastic syndrome

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To the editor,

Myelodysplastic syndrome (MDS) is a clonal disorder of hematopoietic stem cell with maturation defect resulting in ineffective hematopoiesis. A recent mouse study showed that dysfunction of telomere drives MDS [1], which indicates that short telomeres might contribute pathogenesis of MDS. When telomeres become extremely short and sufficient loading of the shelterin complex is not possible, cells undergo senescence and induce cell cycle arrest [2].

Several studies have shown that patients with MDS have a shorter telomere length (TL) than healthy

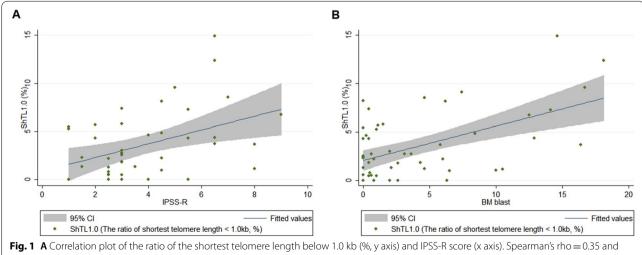


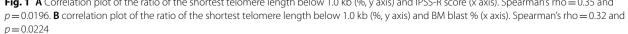
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controls. However, these studies used conventional telomere restriction fragment (TRF) [3] and interphase quantitative fluorescence in situ hybridization (Q-FISH) [4] method, which provide average TL or relative TL. Several methods to measure the absolute length of short telomere such as STELA (Single telomere length analysis) [5], Universal-STELA [6], and TeSLA (Telomere shortest length assay) [7] have been developed. The most recent method to measure short TL, TeSLA was designed to detect TL up to 18 kb of whole chromosomes [7]. Furthermore, Telomeres below 1.6 kb cannot be visualized by other methods except TeSLA [7].

Here, we investigated TL profile including the proportion of cells with critically short TL in bone marrow (BM) nucleated cells from 52 MDS patients by modified TeSLA with the internally labeled biotin probe. To assess critically short TL as a potential prognostic factor, the association of short telomeres with prognostic risk and clinical outcomes was evallated (Supplementary materials and methods).

In this study, a total of 52 patients were included. Further characteristics of patients are described in Supplementary Table 2. A median 177 telomere bands (range 75 – 262) were measured for each single patient's sample. The average TL was 3.18 kb (range, 2.2 - 4.34), which was remarkably shorted than those of healthy populations (5 – 15 kb) previously reported [8, 9]. The shortest TL was 0.787 kb (Supplementary table 3). Our results clearly demonstrated the profiles of shortest telomeres in MDS considering the fact that TL below 1.6 kb was not visualized by other methods except TeSLA [7]. Further details of TL are described in supplementary Table 3. The average TL was not different among MDS (3.32 kb in MDS-SLD, 3.10 kb in MDS-RS-SLD, 3.30 kb in MDS-RS-MLD, 3.18 kb in MDS-MLD, 3.16 kb in MDS-EB1, 3.02 kb in MDS-EB2, 3.43 kb in MDS-U, p=0.71). In contrast, the shortest TL and the ratio of shortest telomere below 1.0 kb (ShTL1.0) showed trends towards positive correlations to MDS subtypes. ShTL1.0 was increased in higher risk MDS group (2.39% in lower risk MDS group and 5.20% in higher risk MDS group, p = 0.0227). The scores of IPSS-R and ShTL1.0 was linearly correlated (spearman's rho=0.35 and p=0.0196) (Fig. 1A). ShTL1.0 showed a positive correlation with blast percentage in BM (spearman's rho=0.32 and p=0.0224) (Fig. 1B). Although ShTL1.0 s according to the presence of thrombocytopenia showed a trend toward a significant difference, we could not find differences of ShTL1.0 values according to the presence of cytopenia or dysplasia (Supplementary Fig. 3). We further investigated if coefficient of variation (CV) of TL was correlated with IPSS-R or BM blasts. The CV of the TL was not significantly correlated with either (Supplementary Fig. 4). MDS patients with a shortest $TL \ge 0.787$ kb at the time of diagnosis showed better OS and a trend toward better PFS compared to patients with a shortest TL < 0.787 kb in univariate analyses (HR = 0.23 and 0.44, p = 0.011 and 0.064, respectively) (Fig. 2). Multivariate analyses showed that the shortest TL was an independent prognostic factor for PFS and OS both (Supplementary tables 4 and 5). Interestingly, when we analyzed the impact of age on OS according to the cut-off of 60 years, survival difference was consistent in a subgroup of elderly patients $(\geq 60$ years, p = 0.028), showed trends towards differences according to the shortest TL for PFS in young patients (< 60 years, p = 0.099) and OS in elderly patients $(\geq 60 \text{ years}, p = 0.084)$ (Supplementary Fig. 5).





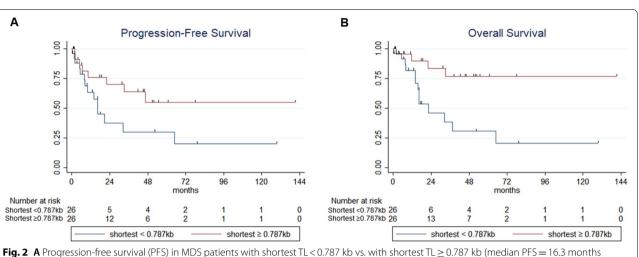


Fig. 2 A Progression-free survival (PFS) in MDS patients with shortest TL < 0.787 kb vs. with shortest TL \ge 0.787 kb (median PFS = 16.3 months vs. not reached [NR], p = 0.0577) and **B** Overall survival in MDS patients with shortest TL < 0.787 kb vs. with shortest TL \ge 0.787 kb (median OS = 22.43 months, vs. NR, p = 0.0056)

Our results suggest that cells with critically short telomeres expand during progression of MDS. The proportion of cells with critically short TL in MDS was a robust indicator to predict poor OS. Our study has a few limitations from its retrospective nature and a small number of patients enrolled into this study. Further, analyses of germline mutations of TERT [10] was not performed in this study. However, our findings on the importance of the presence and abundance of extremely short telomere in the prognosis of MDS can be incorporated into the current prognostic scoring system of the IPSS-R. A large prospective study is needed to establish a new prognostic scoring system in which TL parameters are incorporated as prognostic factors.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s40364-022-00426-9.

Additional file 1.

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Authors' contributions

DS Lee diagnosed MDS through interpretations of bone marrow slides of the patients. DY Shin and SS Yoon treated the patients included in this study. DS Lee designed and initiated this study. KM Lim performed TeSLA experiment. DY Shin and KM Lim analyzed experimental and clinical data of patients and wrote the manuscript. HS Park helped to interpret bone marrow findings and prepared the patients' samples. S Kwon provided a critical help through scientific discussion and interpretation. SS Yoon and DS Lee supervised the study. All authors reviewed the manuscript. The author(s) read and approved the final manuscript.

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Availability of data and materials

Data is available upon the request to corresponding author (Dong-Soon Lee, e-mail to soonlee@snu.ac.kr).

Declarations

Ethics approval and consent to participate

This study was approved by the institutional review board of Seoul National University Hospital (2006–179-1135). Informed consent of the use of bone marrow derived samples for research purposed from patients.

Consent for publication

Institutional review board of Seoul National University Hospital approved this study and publication. All authors agreed the final manuscript for publication.

Competing interests

All authors declared no competing interests.

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References

- Colla S, Ong DS, Ogoti Y, Marchesini M, Mistry NA, Clise-Dwyer K, et al. Telomere dysfunction drives aberrant hematopoietic differentiation and myelodysplastic syndrome. Cancer Cell. 2015;27(5):644–57.
- 2. Maciejowski J, de Lange T. Telomeres in cancer: tumour suppression and genome instability. Nat Rev Mol Cell Biol. 2017;18(3):175–86.
- Ohyashiki JH, Iwama H, Yahata N, Ando K, Hayashi S, Shay JW, et al. Telomere stability is frequently impaired in high-risk groups of patients with myelodysplastic syndromes. Clin Cancer Res. 1999;5(5):1155–60.
- Hwang SM, Kim SY, Kim JA, Park HS, Park SN, Im K, et al. Short telomere length and its correlation with gene mutations in myelodysplastic syndrome. J Hematol Oncol. 2016;9(1):62.
- Baird DM, Rowson J, Wynford-Thomas D, Kipling D. Extensive allelic variation and ultrashort telomeres in senescent human cells. Nat Genet. 2003;33(2):203–7.
- Bendix L, Horn PB, Jensen UB, Rubelj I, Kolvraa S. The load of short telomeres, estimated by a new method, Universal STELA, correlates with number of senescent cells. Aging Cell. 2010;9(3):383–97.
- Lai TP, Zhang N, Noh J, Mender I, Tedone E, Huang E, et al. A method for measuring the distribution of the shortest telomeres in cells and tissues. Nat Commun. 2017;8(1):1356.
- Canela A, Vera E, Klatt P, Blasco MA. High-throughput telomere length quantification by FISH and its application to human population studies. Proc Natl Acad Sci U S A. 2007;104(13):5300–5.
- Kong CM, Lee XW, Wang X. Telomere shortening in human diseases. FEBS J. 2013;280(14):3180–93.
- Reilly CR, Myllymaki M, Redd R, Padmanaban S, Karunakaran D, Tesmer V, et al. The clinical and functional effects of TERT variants in myelodysplastic syndrome. Blood. 2021;138(10):898–911.

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